

YT MicroPlate™

A1 Water	A2 Acetic Acid	A3 Formic Acid	A4 Propionic Acid	A5 Succinic Acid	A6 Succinic Acid Mono-Methyl Ester	A7 L-Aspartic Acid	A8 L-Glutamic Acid	A9 L- Proline	A10 D-Gluconic Acid	A11 Dextrin	A12 Inulin
B1 D-Cellobiose	B2 Gentiobiose	B3 Maltose	B4 Maltotriose	B5 D-Melezitose	B6 D-Melibiose	B7 Palatinose	B8 D-Raffinose	B9 Stachyose	B10 Sucrose	B11 D-Trehalose	B12 Turanose
C1 N-Acetyl-D-Glucosamine	C2 α-D-Glucose	C3 D-Galactose	C4 D-Psicose	C5 L-Sorbose	C6 Salicin	C7 D-Mannitol	C8 D-Sorbitol	C9 D-Arabitol	C10 Xylitol	C11 Glycerol	C12 Tween 80
D1 Water	D2 Fumaric Acid	D3 L-Malic Acid	D4 Succinic Acid Mono-Methyl Ester	D5 Bromo-Succinic Acid	D6 L-Glutamic Acid	D7 γ-Amino-Butyric Acid	D8 α-Keto-Glutaric Acid	D9 2- Keto-D-Gluconic Acid	D10 D-Gluconic Acid	D11 Dextrin	D12 Inulin
E1 D-Cellobiose	E2 Gentiobiose	E3 Maltose	E4 Maltotriose	E5 D-Melezitose	E6 D-Melibiose	E7 Palatinose	E8 D-Raffinose	E9 Stachyose	E10 Sucrose	E11 D-Trehalose	E12 Turanose
F1 N-Acetyl-D-Glucosamine	F2 D-Glucosamine	F3 α-D-Glucose	F4 D-Galactose	F5 D-Psicose	F6 L-Rhamnose	F7 L-Sorbose	F8 α-Methyl-D-Glucoside	F9 β- Methyl-D-Glucoside	F10 Amygdalin	F11 Arbutin	F12 Salicin
G1 Maltitol	G2 D-Mannitol	G3 D-Sorbitol	G4 Adonitol	G5 D-Arabitol	G6 Xylitol	G7 i-Erythritol	G8 Glycerol	G9 Tween 80	G10 L-Arabinose	G11 D-Arabinose	G12 D-Ribose
H1 D-Xylose	H2 Succinic Acid Mono-Methyl Ester plus D-Xylose	H3 N-Acetyl-L-Glutamic Acid plus D-Xylose	H4 Quinic Acid plus D-Xylose	H5 D-Glucuronic Acid plus D-Xylose	H6 Dextrin plus D-Xylose	H7 α-D-Lactose plus D-Xylose	H8 D-Melibiose plus D-Xylose	H9 D-Galactose plus D-Xylose	H10 m-Inositol plus D-Xylose	H11 1,2-Propanediol plus D-Xylose	H12 Acetoin plus D-Xylose

FIGURE 1. Carbon Sources in YT MicroPlate



Oxidation Tests



Assimilation Tests

INTRODUCTION

The YT MicroPlate™ provides a broad capability for identification and characterization of yeast strains, including both human isolates and environmental species. Yeast are of particular importance in the food industry, both in food production and in food spoilage. They are also important in human health both as normal flora (e.g. in the gastrointestinal tract) and as occasional pathogens. There has been a renewed interest in the use of yeast as “probiotics” to beneficially influence the ecology of the digestive tract and the ecology of plant surfaces.

The unique physiological properties of yeast have made them relatively difficult to test and identify. Yeast tend to thrive in low pH and high sugar environments. Most species have a slower growth rate and metabolism as compared to common bacteria.

YT MICROPLATE

The Biolog YT MicroPlate™ (Figure 1) is designed for identification and characterization of a very wide range of Yeasts. Biolog’s MicroPlates and databases were first introduced in 1989, employing a novel, patented redox chemistry. This chemistry, based on reduction of tetrazolium, responds to the process of metabolism (i.e. respiration) rather than to metabolic by-products (e.g. acid). Biolog’s chemistry works as a universal reporter of metabolism and simplifies the testing process as color developing chemicals do not need to be added. Since the YT MicroPlate™ measures both metabolic reactions as well as turbidity growth to produce identifications, it provides superior capability for all types of yeasts organisms. The database for the YT Microplate™ is now over 267 species. It is by far the largest kit based identification database available.

PROCEDURE FOR USING YT MICROPLATES

The Biolog System makes identifying yeast nearly as easy to identify as bacteria. The testing protocol is a very simple one:

- 1) The strain of interest is cultured on a special agar medium, BUY™ Agar (available for Biolog either as dry powder – Catalog No. 70005 or already prepared in Petri plates – Catalog No. 71005)
- 2) Cells are removed from the surface of the agar with a sterile swab and suspended in sterile water at the specified density.
- 3) 100 µl of the cell suspension is inoculated into each of the 96 wells of the Biolog YT MicroPlate (carbon sources shown schematically above),
- 4) The MicroPlate is incubated at 26-28°C for 24, 48 or 72 hours until a sufficient metabolic pattern is formed.
- 5) For identification the MicroPlates are read with the MicroStation™ or the OmniLog™ Plus system and compared to the YT database. (Biolog Catalog No. 22605D)

Some yeast species are inhibited by the tetrazolium violet redox used in Biolog MicroPlates, so the YT MicroPlate is configured with both metabolism test and turbidity tests. The first 3 rows of the panel (rows A-C) contain carbon source metabolism tests using tetrazolium violet as a colorimetric indicator. The next five rows of the panel (rows D-H) contain carbon source turbidity tests. Results

from this test are scored turbidimetrically. The last row of the panel (row H) has wells that contain 2 carbon sources. These wells test for the co-utilization of various carbon sources with D-Xylose.

For manual characterization of yeast strains, reactions may be read by eye. Metabolism test rows A-C should be read against a white background and turbidity tests in rows D-H should be read against a black background. Depending on the strain, some reactions may be faint and difficult to read by eye.

For species identification, the YT MicroPlate must be read with the Biolog MicroStation Reader. A list of the 267 species of yeast identified by the Biolog System is shown on the back of this sheet.

CONTACT INFORMATION

The Biolog Microbial Identification/Characterization System will be an invaluable addition to your microbiology laboratory. Incidentally, our FF MicroPlate also has a subset of 76 yeast species for identification.

For more details, contact us using the information below:

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